

PRODUCT DESCRIPTION

Arachidonic Acid Reagent is a lyophilized preparation of the Sodium Salt of Arachidonic Acid. It is an essential fatty acid present in the granules of platelets and on the platelet membrane. It is processed in multiple steps and converted to Thromboxane A₂ (TXA₂). Arachidonic Acid Reagent induces platelet activation and aggregation.

Arachidonic Acid Reagent has been optimized for use with Light Transmission Aggregometers. It may also be used with other turbidometric or impedance analyzers, and flow cytometers.

INTENDED PURPOSE

Arachidonic Acid Reagent (Sodium Arachidonate) is for routine use in demonstrating thromboxane A₂ activation response in Platelet Rich Plasma (PRP) test samples.

DETECTION / MEASUREMENT

Arachidonic Acid Reagent is used, in conjunction with other diluents and control samples, to measure changes of the light transmission in a Platelet Rich Plasma (PRP) test sample.

PRODUCT FUNCTION

Arachidonic Acid Reagent provides insight into different aspects of platelet function / quality. This Reagent aids in assessing various acquired and inherited platelet disorders or the efficacy of anti-platelet therapies.

SPECIFIC INFORMATION PROVIDED

Arachidonic Acid Reagent is not intended for the detection of a specific disorder, condition, or risk factor.

Arachidonic Acid Reagent initiates platelet activation and aggregation through the arachidonic acid pathway. Upon binding to platelet surface receptors, arachidonic acid undergoes enzymatic conversion to Thromboxane A₂ (TXA₂), facilitating intracellular signaling cascades. This prompts rapid changes in platelet shape and calcium ion release, crucial for stable aggregation. Observing platelet aggregation in response to Arachidonic Acid Reagent allows clinicians to assess and evaluate platelet function / quality, abnormalities, and anti-platelet therapies. Arachidonic Acid Reagent's induction of secondary mediators like Thromboxane A₂ (TXA₂) amplifies platelet activation.

AUTOMATION

Arachidonic Acid Reagent is intended for use in semi-automated and automated Light Transmission Platelet Aggregometers. This Reagent may also be used with other turbidometric or impedance analyzers, and flow cytometers.

QUALITY / QUANTITY

There are no primary standards for the Arachidonic Acid Reagent. The responses to this reagent are concentration dependent. A known normal donor should be tested with each new lot of Arachidonic Acid Reagent. Standards organizations classify Arachidonic Acid induced platelet aggregation as semi-quantitative or semi-qualitative.

Arachidonic Acid Reagent comes packaged as 3 x 0.5 mL vials. The working concentration of Arachidonic Acid is 5 mg / mL.

SPECIMEN TYPE

The test specimen is prepared from sodium citrate anti-coagulated whole blood. The test sample is Platelet Rich Plasma (PRP). The test blank is Platelet Poor Plasma (PPP).

Arachidonic Acid Reagent may be used with human or animal Platelet Rich Plasma (PRP) for routine platelet aggregation tests. Results are based on the concentration, extent, and rate of aggregation compared to a Platelet Poor Plasma (PPP) blank.

TESTING POPULATION

- Human: The prevalence of platelet disorders is global and may vary by race, ethnicity, blood type, and other factors. The incidence is variable.
- Anti-Platelet Drugs: The prevalence of abnormal Arachidonic Acid Reagent aggregation, contingent on estimated Aspirin usage, reaches up to one third of the population. Both Clopidogrel and the combination of Clopidogrel with Aspirin can influence Arachidonic Acid-induced platelet aggregation. The incidence is variable.
- Inherited Platelet Disorders: The prevalence and incidence are variable. There are 60 types of inherited platelet disorders that affect approximately 0.3% of the population. Certain inherited platelet defects, such as Glanzmann's Thrombasthenia and Storage Pool Disease, show no response to Arachidonic Acid Reagent.

- Animal: The prevalence and incidence are species dependent.

IN VITRO DIAGNOSTIC

Arachidonic Acid Reagent is an in vitro diagnostic reagent intended for Professional Laboratory Use Only. This Reagent is not intended for injection or ingestion.

INTENDED USER

Arachidonic Acid Reagent is intended for Professional Laboratory Use by qualified personnel.

TEST PRINCIPLE

When introduced to a stirred, 37°C Platelet Rich Plasma (PRP) test sample, exogenous Reagents such as ADP, Arachidonic Acid, Collagen, Epinephrine, and Ristocetin stimulate platelets, prompting them to undergo shape change and aggregate. This initial aggregation is called primary aggregation and is reversible. However, normal platelets possess the ability to release endogenous ADP from their granules, leading to a secondary, irreversible wave of aggregation. The Light Transmission Platelet Aggregometer effectively captures these changes by displaying parameters such as lag phase, shape change, and the rate and extent of aggregation over a predetermined testing period.

CALIBRATORS AND CONTROLS

There are no calibrators or controls required for Arachidonic Acid Reagent. A known donor sample should be tested with each lot of Arachidonic Acid Reagent. Responses are concentration dependent.

REAGENT LIMITATIONS

Arachidonic Acid Reagent will perform as specified when the Instructions for Use are followed. The reagents must be used prior to the expiration date printed on each vial.

REAGENTS PROVIDED

REF 101297: 3 vials of Arachidonic Acid Reagent (0.5 mL)

REAGENTS AND MATERIALS REQUIRED BUT NOT PROVIDED

- Purified Water (Distilled, Deionized, Reagent Grade), pH 5.3 – 7.2 for reconstitution
- TRIS Buffered Saline (TBS) or 0.85% physiological saline for dilutions




 **NOTE: USING BLOOD BANK SALINE WILL CAUSE ERRONEOUS RESULTS.**

MATERIALS AND ACCESSORIES

- Platelet Aggregometer (*Follow the Manufacturer's Instructions for Use*)
- Centrifuge
- Electronic Pipette
- Pipette Tips ②
- Aggregometer Test Tubes (*Siliconized*) ②
- Aggregometer Stir Bars (*Plastic Coated*) ②
- Plastic Sample Tubes and Caps (*for Dilutions*) ②

 **NOTE: DISPOSABLE ITEMS SUCH AS TEST TUBES, STIR BARS, SAMPLE TUBES, AND CAPS ARE FOR ONE TIME USE ONLY**









STORAGE AND STABILITY

-  Arachidonic Acid Reagent does not require temperature protection during shipment.
-  Upon receipt, store Arachidonic Acid Reagent at 2 – 8° C in its original packaging.
-  Reconstituted Arachidonic Acid Reagent is stable for 24 hours when stored in its tightly capped, original containers at 2 – 8° C.

STERILITY

 Arachidonic Acid Reagent is not a sterile product. Be careful not to contaminate the product when pipetting the reconstituted or aliquoted reagents.

WARNINGS AND PRECAUTIONS

-  Wear PPE in accordance with laboratory policies and practices when handling Arachidonic Acid Reagent.
 -  Follow standard precautions when preparing test specimens and samples.
 -  Handle Arachidonic Acid Reagent with care to avoid contamination during use.
 -  Avoid reagent evaporation by limiting air – liquid exchange surfaces.
 -  To ensure optimum test results, a known donor control sample should be run consecutively, without interruption.
 -  To preserve reagent stability, store remaining reagent in its tightly capped, original containers.
 -  Dispose of post-test materials in accordance with applicable regulations and laboratory policies.
-  **NOTE TO USER: ANY SERIOUS INCIDENT THAT OCCURS IN RELATION TO THIS PRODUCT SHALL BE REPORTED TO THE MANUFACTURER AND THE COMPETENT AUTHORITY OF THE MEMBER STATE IN WHICH THE USER AND / OR PATIENT ARE ESTABLISHED.**

INFECTIOUS MATERIAL STATUS

Arachidonic Acid Reagent does not contain any infectious materials. Test specimens and samples must be considered infectious and should be handled as if capable of transmitting infection. After testing, test specimens and samples must be disposed of in compliance with applicable regulations and laboratory policies.

SPECIAL FACILITIES

Arachidonic Acid Reagent does not require the use of special facilities within a laboratory environment.

PREPARATION FOR USE

-  **NOTE: ARACHIDONIC ACID REAGENT MUST BE AT ROOM TEMPERATURE (15 – 28° C) PRIOR TO RECONSTITUTION. STORED REAGENTS MUST BE BROUGHT TO ROOM TEMPERATURE PRIOR TO USE.**

RECONSTITUTION

The working concentration of the reconstituted Arachidonic Acid Reagent is 5 mg / mL. All final concentrations are based on adding 25 µL of Arachidonic Acid Reagent to a 225 µL Platelet Rich Plasma (PRP) test sample.

- Reconstitute Arachidonic Acid Reagent with 0.5 mL of Purified Water.
- Invert gently to mix.

-  **NOTE: ARACHIDONIC ACID REAGENT MAY APPEAR CLOUDY BUT WILL BECOME CLEAR TO PALE YELLOW WITHIN A FEW MINUTES.**

- Reconstituted Arachidonic Acid Reagent should be kept capped prior to use.


PATIENT PREPARATION

Patients should refrain from taking aspirin or using aspirin-containing medications and products, as well as other medications, supplements, or energy drinks known to affect platelet function for 7 – 10 days prior to specimen collection. Ingestion of fatty foods, dairy products, and smoking should be avoided for 12 hours before specimen collection.

-  **NOTE: CONSULTATION WITH A PHYSICIAN IS REQUIRED PRIOR TO MAKING ANY MEDICATION CHANGES.**

SPECIMEN COLLECTION

The specimen should be collected with care to avoid stasis, hemolysis, contamination by tissue fluid and exposure to glass. Specimens must be kept at room temperature. Release the tourniquet as soon as blood begins to flow into the collection device.


-  **PRACTICE STANDARD PRECAUTIONS THROUGHOUT THE SPECIMEN COLLECTION, SAMPLE PREPARATION, AND ANALYTICAL PROCESSES. DISPOSE OF SHARPS AND BIOHAZARDOUS WASTE IN ACCORDANCE WITH APPLICABLE REGULATIONS AND LABORATORY POLICIES.**

Evacuated Specimen Collection Technique

- Use a 21g or 23g winged needle collection set for specimen collection
- Draw blood into plastic evacuated specimen collection tubes containing 3.2% (0.11 M) sodium citrate anti-coagulant
- Gently mix the specimen collection tube 4 - 5 times by inversion
- Write collection time on the specimen label
- Maintain specimen collection tubes at room temperature
- Remix specimen collection tubes prior to centrifugation

Syringe Collection Technique

- Use a 21g or 23g winged needle collection set for the venipuncture
- Draw 9.0 mL of blood into a plastic syringe, avoiding excess suction
- Clamp the winged needle tubing and disconnect the syringe
- Immediately and gently dispense the blood specimen into a plastic (polypropylene) tube containing 1.0 mL of 0.11 M sodium citrate anti-coagulant. The blood to anticoagulant ratio is 9 parts blood to 1 part anti-coagulant
- Cap the plastic tube
- Gently mix the specimen collection tube 4 - 5 times by inversion
- Write collection time on the specimen label
- Maintain specimen collection tubes at room temperature
- Remix specimen collection tubes prior to centrifugation

-  **NOTE: WHEN THE PATIENT'S HEMATOCRIT IS LESS THAN 30% OR GREATER THAN 55%, THE BLOOD TO ANTI-COAGULANT RATIO MUST BE ADJUSTED. BLUE TOP EVACUATED SPECIMEN COLLECTION TUBES MUST CONTAIN 3.2% (0.11 M) SODIUM CITRATE ANTICOAGULANT, WHICH IS THE RECOMMENDED CONCENTRATION FOR PLATELET FUNCTION STUDIES.**

SAMPLE PREPARATION

Platelet Rich Plasma (PRP)

- Centrifuge the anti-coagulated blood at 150 x g for 10 minutes at room temperature
- Examine the plasma layer for red cells
- If red cells are present, re-centrifuge for an additional 5 minutes
- Use a Pipette to transfer the PRP to a plastic container labeled PRP
- Remove the PRP from a point just below the middle of the PRP volume for consistent platelet count (**THE TOP OF THE VOLUME HAS A LOWER PLATELET COUNT AND THE BOTTOM IS MORE CONCENTRATED**)
- Cap the container
- Allow the container to stand at room temperature

Platelet Poor Plasma (PPP)


- Centrifuge the remaining PRP blood specimen at 2500 x g for 20 minutes
- Use a Pipette to transfer the PPP to a plastic container labeled PPP
- Cap the container
- Allow the container to stand at room temperature

ASSAY PROCEDURE

Routine Aggregation Procedure

-  **NOTE: THIS IS A GENERAL PROCEDURE. FOLLOW THE INSTRUCTIONS FOR USE PROVIDED BY THE MANUFACTURER OF THE AGGREGOMETER IN USE.**

Prepare a Blank for Each Patient

-  **NOTE: EACH PATIENT MUST HAVE THEIR OWN BLANK. ONE PATIENT'S BLANK CANNOT BE USED FOR ANY OTHER PATIENT. THE PATIENT'S BLANK MUST BE PREPARED FROM THE PATIENT'S PLATELET POOR PLASMA (PPP) SPECIMEN. IF THE SAME PATIENT IS BEING TESTED ON MULTIPLE TEST WELLS, THE SAME PATIENT'S BLANK MAY BE USED FOR THOSE TEST WELLS.**

- Label a test tube with the letter "B", test well #, and patient ID to identify the Blank
- Pipette 250 µL of Platelet Poor Plasma (PPP) into the test tube (**DO NOT ADD A STIR BAR**)
- Place Blank aside for later use
- Repeat the steps above for each patient

Prepare Samples

- Label one to eight new test tubes with each patient ID and test well #
- Place the labeled test tubes into the correct well # 1 - 8 of the stirred sample incubation wells
- Add a stir bar to each test tube
- Pipette 225 µL of Platelet Rich Plasma (PRP) sample into each test tube in the stirred sample incubation wells (**MAKE SURE THERE ARE NO BUBBLES**)
- Select the on-screen timer for each stirred sample incubation well in use and the warming countdown will start
- The samples will incubate at 37° C for the pre-set time
- Set the 100% baseline (Blank)
- Place the appropriate previously prepared patient's Blank test tube into test well # 1
- Select BLANK to activate the test well
- The BLANK button will change to START
- Repeat the steps above for each test well being used for testing

Begin Testing

- Once the countdown timer reaches 0:00, press the timer button to stop each stirred sample incubation well
- Transfer the test tube in the stirred sample incubation well # 1 to test well # 1
- Repeat the step above for each test well, making sure all test tubes remain with their corresponding well #'s during transfer

FIGURE 1: ARACHIDONIC ACID NORMAL AGGREGATION

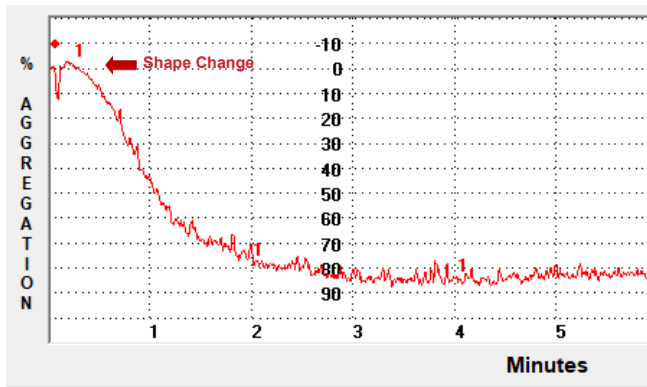
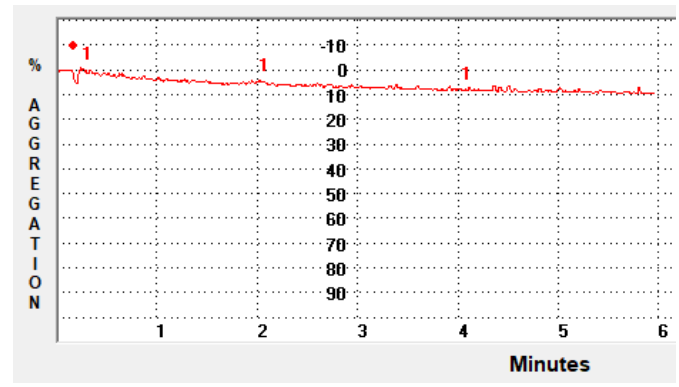


FIGURE 2: ARACHIDONIC ACID ABNORMAL RESPONSE (Aspirin Effect)



- Close the pipette guides
- Select START for test well # 1
- Pipette 25 μ L of reagent directly into the Platelet Rich Plasma (PRP) test tube in test well # 1 (**DO NOT ALLOW REAGENT TO RUN DOWN THE WALL OF THE TEST TUBE OR PERMIT PIPETTE TIP TO BREAK THE SURFACE OF THE SAMPLE**)
- Select INJECT for test well # 1
- Repeat the steps above for each test well being used for testing
- The test will now run for the pre-set time (**OTHER MANUFACTURER'S TEST PROCEDURES MAY SPECIFY DIFFERENT TIMES OR VOLUMES**)

NOTE: USE A KNOWN DONOR AS A CONTROL SAMPLE. EACH LABORATORY SHOULD ESTABLISH AND VALIDATE ITS OWN TEST PROTOCOL AND VERIFY THE RESULTING PERFORMANCE OF ITS TEST SYSTEM (REAGENTS, INSTRUMENT, AND TEST PROTOCOL).

QUALITY CONTROL

For platelet aggregation studies, a known donor should be tested in the same manner as the patient to ensure test system performance and consistency. A new control should be included with each test series, and preferably with each new reagent lot or after instrument maintenance. Each laboratory must define its acceptable ranges for its patient population and verify the expected performance of the test system.

RESULTS

Typical aggregation patterns induced by Arachidonic Acid Reagent are illustrated in Figures 1 and 2. These patterns provide a comprehensive view of how the reagent interacts with Platelet Rich Plasma (PRP) under different conditions.

Ingestion of a single 600 mg dose of Aspirin has a significant impact on platelet aggregation, resulting in the absence of Arachidonic Acid-induced aggregation for up to 5 days, as demonstrated in Figure 1. This absence indicates that Aspirin effectively inhibits the aggregation response, which is crucial for understanding its anticoagulant properties.

Furthermore, a prolonged response time can be observed for up to 8 days following Aspirin ingestion, as depicted in Figure 2. This prolonged response time refers to the delay from the addition of Arachidonic Acid Reagent to the onset of aggregation, highlighting the extended effect of Aspirin on platelet function.

Spike marks in the figures indicate the points at which the reagent was added, providing clear reference points for the timing of reagent introduction and its effects on the aggregation process.

TABLE 1: ARACHIDONIC ACID RESULTS OBSERVED IN PLATELET FUNCTION DEFECTS

| DEFECT | ARACHIDONIC ACID REAGENT |
|--------------------------|--------------------------|
| ASPIRIN-LIKE | ↓ or N |
| THROMBASTHENIA | ↓↓ ↓↓ |
| STORAGE POOL DISEASE | ↓ |
| VON WILLEBRAND SYNDROME | N |
| BERNARD-SOULIER SYNDROME | N |

↓ = Reduced Aggregation Resulting From a Decrease or Absence of Secondary Wave
 ↓↓ = Reduced Aggregation Resulting From a Decrease or Absence of Primary and Secondary Wave
 N = Normal Response

EXPECTED VALUES

Each laboratory must establish its own expected ranges and performance characteristics for this reagent at the concentrations used to induce platelet aggregation. These ranges should be determined using the laboratory's specific instrumentation, procedures, reference intervals, and patient population.

Published literature reports that Arachidonic Acid Reagent typically produces a Final Aggregation response in the range of 61–93% and a Lag Phase of \geq 25 seconds, under standard test conditions. This literature-based range is provided as general information only; laboratories must verify and establish their own expected ranges before clinical use.

LIMITATIONS

In Light Transmission Aggregometry, the presence of red blood cells in the PRP will cause the observed aggregation to be reduced. The presence of platelets in the PPP will cause final aggregation to be increased. Spurious results may occur if the PRP platelet count is less than 75,000 platelets / μ m. PRP platelet counts can only be performed using the hemocytometer method. Compromised samples must be rejected. If the results are abnormal, the test should be repeated on another occasion. Each laboratory must establish reference ranges tailored to the population it serves, and the specific reagent concentrations used.

ANALYTICAL PERFORMANCE

Platelet aggregation, induced by commonly used reagents like Arachidonic Acid Reagent, is a non-linear test system. Responses are based on the difference between the patient's Platelet Rich Plasma (PRP) and Platelet Poor Plasma (PPP) light transmission and therefore, results are unique to that patient. Certain parameters are more prone to non-linearity than others. These include lag phase, primary slope, secondary slope, biphasic response and disaggregation. The non-linearity is caused by many factors such as the reaction chemistry and instrumentation. Platelet aggregation displays the response rate or activity and does not quantify the reactants or their concentrations.

In platelet aggregation, accuracy is a relative parameter and is dependent on the test system. The limitations of platelet aggregation make it difficult to provide typical precision or reproducibility ranges.

The variability in linearity, precision and reproducibility of results in Arachidonic Acid Reagent-based test systems is acknowledged by multiple standards organizations. The commonly accepted CV is \pm 15%.

| | |
|---|-----------------------|
| Test to Test Reproducibility: | less than \pm 7.5% |
| Instrument to Instrument Reproducibility: | less than \pm 15.0% |
| Reagent Lot to Lot Variability: | less than \pm 10.5% |
| Laboratory to Laboratory (System to System) | less than \pm 12.5% |

REFERENCES

- Allain JP, Cooper HA, Wagner RH, Brinkhous KM. Platelets fixed with paraformaldehyde: a new reagent for assay of von Willebrand factor and platelet aggregating factor. J Lab Clin Med. 1975 Feb;85(2):318-28.
- Angiolillo DJ, Ueno M, Goto S. Basic principles of platelet biology and clinical implications. Circ J. 2010 Apr;74(4):597-607.
- Born GV, Cross MJ. The Aggregation of Blood Platelets. J Physiol. 1963 Aug; 168(1):178-95.
- Brinkhous KM, Read MS. Preservation of platelet receptors for platelet aggregating factor/von Willebrand factor by air drying, freezing, or lyophilization: new stable platelet preparations for von Willebrand factor assays. Thromb Res. 1978 Oct;13(4):591-7.
- Bye A, Lewis Y, O'Grady J. Effect of a single oral dose of aspirin on the platelet aggregation response to arachidonic acid. Br J Clin Pharmacol. 1979 Mar; 7(3):283-6.
- Cattaneo M, Cerletti C, Harrison P, Hayward CP, Kenny D, Nugent D, Nurden P, Rao AK, Schmaier AH, Watson SP, Lussana F, Pugliano MT, Michelson AD. Recommendations for the Standardization of Light Transmission Aggregometry: A Consensus of the Working Party from the Platelet Physiology Subcommittee

of SSC/ISTH. J Thromb Haemost. 2013 Apr 10.

- CLSI. Procedures for the Handling and Processing of Blood Specimens for Common Laboratory Tests; Approved Guideline—Fourth Edition. CLSI document H18-A4. Wayne, PA: Clinical and Laboratory Standards Institute; 2010.
- CLSI. Protection of Laboratory Workers from Occupationally Acquired Infections, Approved Guideline - Fourth Edition. CLSI document M29-A4. Wayne, PA: Clinical and Laboratory Standards Institute; 2010.
- CLSI. Platelet Function Testing by Aggregometry, Approved Guideline - Fourth Edition. CLSI document H58-A. Wayne, PA: Clinical and Laboratory Standards Institute; 2008.
- CLSI. Collection, Transport and Processing for Plasma Based Coagulation Assays and Molecular Hemostasis Assays, Approved Guideline - Fifth Edition. CLSI document H21-A5. Wayne, PA: Clinical and Laboratory Standards Institute; 2008.
- CLSI. Clinical Laboratory Safety, Approved Guideline - Third Edition. CLSI document GP17-A3. Wayne, PA: Clinical and Laboratory Standards Institute; 2012.
- Day HJ, Holmsen H. Laboratory tests of platelet function. Ann Clin Lab Sci (1971). 1972 Jan-Feb; 2(1):63-74.
- Day HJ, Rao AK. Evaluation of platelet function. Semin Hematol. 1986 Apr;23(2):89-101.
- Eichelberger, JW. Kinetic (Slope) Measurement of Platelet Aggregation. Bio/Data Corporation, Horsham, PA; 1984.
- Favaloro EJ, Gosselin RC, Pasalic L, Lippi G. Post-analytical issues in hemostasis and thrombosis testing: An update. In EJJ, RCG, editors, Hemostasis and Thrombosis: Methods and Protocols. 2nd ed. New York: Humana Press. 2023. p. 787-811. (Methods in Molecular Biology).
- Federici AB, Lee CA, Berntorp EE, Lillicrap D, Montgomery RR. Von Willebrand Disease: Basic and Clinical Aspects. 2011.
- Garner JS. Guideline for isolation precautions in hospitals. The Hospital Infection Control Practices Advisory Committee. Infect Control Hosp Epidemiol. 1996 Jan;17(1):53-80.
- Howard MA, Firkin BG. Ristocetin—a new tool in the investigation of platelet aggregation. Thromb Diath Haemorrh. 1971 Oct 31; 26(2): 362-9.
- Israels SJ, El-Ekiaby M, Quiroga T, Mezzano D. Inherited disorders of platelet function and challenges to diagnosis of mucocutaneous bleeding. Haemophilia. 2010 Jul;16 Suppl 5:152-9.
- Kambayashi J, Shinoki N, Nakamura T, Ariyoshi H, Kawasaki T, Sakon M, Monden M. Prevalence of impaired responsiveness to epinephrine in platelets among Japanese. Thromb Res. 1996 Jan 1;81(1):85-90.
- Levine PH. The effect of thrombocytopenia on the determination of platelet aggregation. Am J Clin Pathol. 1976 Jan;65(1):79-82
- Linnemann B, Schwonberg J, Mani H, Prochnow S, Lindhoff-Last E. Standardization of light transmittance aggregometry for monitoring antiplatelet therapy: an adjustment for platelet count is not necessary. J Thromb Haemost. 2008 Apr;6(4):677-83.
- Marcus AJ, Coleman RW, Hirsh J, Ivarer VJ, Salzman EW. Hemostasis and thrombosis: Basic Principles and Clinical Practice. Vol. 472. Philadelphia: JB Lippincott Company; 1982.
- Michelson, AD. Platelets. Third Edition. Amsterdam: Academic Press; 2013.
- Mills DC, Robb IA, Roberts GC. The release of nucleotides, 5-hydroxytryptamine and enzymes from human blood platelets during aggregation. J Physiol. 1968 Apr;195(3):715-29.
- Moncada S, Vane JR. Arachidonic acid metabolites and the interactions between platelets and blood-vessel walls. N Engl J Med. 1979 May 17;300(20):1142-7.
- NCCLS. Assays of von Willebrand Factor Antigen and Ristocetin Cofactor Activity; Approved Guideline. NCCLS document H51-A. NCCLS, 940 West Valley Road, Suite 1400, Wayne, Pennsylvania 19087-1898 USA, 2002.
- O'Donnell CJ, Larson MG, Feng D, Sutherland PA, Lindpaintner K, Myers RH, D'Agostino RA, Levy D, Tofler GH; Framingham Heart Study. Genetic and environmental contributions to platelet aggregation: the Framingham heart study. Circulation. 2001 Jun 26;103(25):3051-6.
- Owen CA Jr, Bowie EJW, Thompson JH Jr. The Diagnosis of Bleeding Disorders. 2nd ed. Little, Brown, and Company; 1975.
- Palma-Barqueros V, Revilla N, Sánchez A, Zamora Cánovas A, Rodríguez-Alén A, Marín-Quílez A, González-Porras JR, Vicente V, Lozano ML, Bastida JM, Rivera J. Inherited Platelet Disorders: An Updated Overview. Int J Mol Sci. 2021 Apr 26;22(9):4521.
- Siegel JD, Rhinehart E, Jackson M, Chiarello L; Health Care Infection Control Practices Advisory Committee. 2007 Guideline for Isolation Precautions: Preventing Transmission of Infectious Agents in Health Care Settings. Am J Infect Control. 2007 Dec;35(10 Suppl 2):S65-164.
- The Hospital Infection Control Practices Advisory Committee, Centers for disease Control and Prevention, Public Health Service, U.S. Department of Health

and Human Services. Guideline for isolation precautions in hospitals Part II. Recommendations for isolation precautions in hospitals. American Journal of Infection Control. 1996; Vol 24, Issue 1: 32-52.

- Triplett DA, et al. Platelet function: laboratory evaluation and clinical application. Chicago, IL: American Society for Clinical Pathology 1978.
- Weiss HJ. Aspirin and Platelets in Drugs and Hematologic Reactions. New York, NY: Dimittov and Nodine, eds. Grune and Stratton. 1974.
- White, M.M., and Jennings, L.K. Platelet Protocols: Research and Clinical Laboratory Procedures, Academic Press, Inc.; 1999.
- Williams WJ, Beutler E, Erslev AJ, Rundles RW. Hematology. New York, NY: McGraw-Hill. 1977.

SYMBOLS

| | |
|---|---|
|  | Bio-Hazardous |
|  | Catalog Number |
|  | Caution |
|  | CE Marked & Registered Product |
|  | Consult Instructions For Use |
|  | European Union Representative |
|  | In Vitro Diagnostic Device |
|  | Manufacturer |
|  | Must Read |
|  | Non-Sterile |
|  | Single Use Only |
|  | Temperature Limitations |
|  | United Kingdom Marked & Registered Product |
|  | United Kingdom Representative |

REVISION HISTORY

Document No: 101302 Revision: AA, January 2025

- Modified Testing Instructions
- Implemented IVDR Regulatory Requirements
- Reformatted and Reconfigured to Enhance Operator Use

Document No: 101302 Revision: AB, December 2025

- Corrected typographical errors throughout the document, including stability information (24 hours after reconstitution); no changes to product or performance.
- Updated Expected Results section: removed results chart, added literature-based Arachidonic Acid range statement, and clarified that laboratories must establish their own expected ranges.

For a complete product catalog, please visit our website at www.biodatacorp.com or contact our Customer Service Department.

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